

Plant Diagnostic Laboratory

206 Waldron Hall, NDSU, Fargo ND 58102
 Website: www.ag.ndsu.nodak.edu/diaglab
 E-mail: NDSU.PDL@ndsu.edu
 Telephone: 701-231-7854 Fax: 701-231-7851

Lab use only:	
Date In:	
Lab #:	
PDIS #:	
Source:	
Condition:	<input type="checkbox"/> good <input type="checkbox"/> fair <input type="checkbox"/> poor

Information Request Form

Submitted By: _____ Submitted For (Client): _____
 Company: _____ Company: _____
 Address: _____ Address: _____
 City/State/Zip: _____ City/State/Zip: _____
 Phone: _____ Fax: _____ Phone: _____ Fax: _____
 Cell: _____ Cell: _____
 E-mail: _____ E-mail: _____

Extension educator Individual Grower Hort/Turf professional Extension educator Individual Grower Hort/Turf professional
 Consultant Company Rep Other: Consultant Company Rep Other:

Charges: Routine Diagnosis: \$15 ND resident; \$25 non-resident; Culture: \$30 additional; ELISA: \$35. Plant/Insect ID: \$15; Fungus/Mold ID: \$15
 Do you accept additional charges to complete the diagnosis? Yes No

Send samples to: NDSU Plant Diagnostic Lab NDSU Dept 7660 PO Box 6050 Fargo, North Dakota 58108-6050	Services requested: <input type="checkbox"/> Routine Diagnosis <input type="checkbox"/> Plant ID <input type="checkbox"/> Culture/ELISA <input type="checkbox"/> Insect ID <input type="checkbox"/> Special Test <input type="checkbox"/> Fungus ID <input type="checkbox"/> Other:	Results and Billing: Send results to: <input type="checkbox"/> Submitter <input type="checkbox"/> Client Send bill to: <input type="checkbox"/> Submitter <input type="checkbox"/> Client
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Sample collection instructions: See reverse for details.

Host: _____ **Planting Date:** _____ **Symptom development:**
Variety: _____ Date sample collected: _____ Days Weeks Months
 County location: _____ Date sample sent: _____ Occurred in previous years
Turfgrass: Year established: _____ Sod Seed Plugs **Trees/shrubs/ornamentals:** Approx. age: _____ Height: _____
 Number of years at site: _____

Location <input type="checkbox"/> Crop/Field <input type="checkbox"/> Greenhouse <input type="checkbox"/> Golf course <input type="checkbox"/> Lawn/Turf <input type="checkbox"/> Landscape <input type="checkbox"/> Home - interior <input type="checkbox"/> Nursery/Orchard <input type="checkbox"/> Pasture <input type="checkbox"/> Garden <input type="checkbox"/> Other:	Size of Planting _____ Acres _____ # of plants Incidence _____ Acres _____ sq. ft. _____ % of area -- Or -- _____ # of plants _____ % of plants	Symptoms <input type="checkbox"/> Yellowing <input type="checkbox"/> Stunted <input type="checkbox"/> Wilting <input type="checkbox"/> Rot <input type="checkbox"/> Dead areas <input type="checkbox"/> Dieback <input type="checkbox"/> Leaf drop <input type="checkbox"/> Abnormal growth <input type="checkbox"/> Other:	Parts Affected <input type="checkbox"/> Stems/trunk <input type="checkbox"/> Roots <input type="checkbox"/> Leaves <input type="checkbox"/> Flowers <input type="checkbox"/> Fruits/seeds <input type="checkbox"/> Entire plant <input type="checkbox"/> Branches__% <input type="checkbox"/> Other: Distribution on Plant <input type="checkbox"/> Upper canopy <input type="checkbox"/> Lower canopy <input type="checkbox"/> Random	Distribution in Field <input type="checkbox"/> High areas <input type="checkbox"/> Low areas <input type="checkbox"/> Scattered plants <input type="checkbox"/> Groups of plants <input type="checkbox"/> Uniform <input type="checkbox"/> Wet areas <input type="checkbox"/> Sunny spots <input type="checkbox"/> Shady spots <input type="checkbox"/> Edge of planting <input type="checkbox"/> Other:	Field History Soil pH: _____ Soil drainage: <input type="checkbox"/> Good <input type="checkbox"/> Moderate <input type="checkbox"/> Poor Previous crops: Yr 1: _____ Yr 2: _____ Yr 3: _____
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Tillage practices: _____ **Chemical history:** Please provide chemical name, application dates, and rates:
 Fertilizer:
 Seed treatment:
 Herbicide:
 Insecticide:
 Fungicide:

Please describe problem in detail on reverse side

Detailed description of problem and additional information:

How to collect and ship samples:

PLANTS: For general plant problems, try to send several affected plants showing a range of symptoms. Dead plants rarely are informative – avoid sending completely dead plants. Try to send entire plants, when feasible, since some above-ground symptoms can be attributed to a problem with the lower stem or roots. When digging plants, try to keep the roots intact with the soil, as a root ball, to help prevent the sample from drying out. The root ball should be wrapped in damp (not wet) paper towels, and wrapped in a separate plastic bag (tied off at the stem) to prevent soil from coming in contact with leaves. Wrap foliage in DRY paper towels (to absorb moisture and to prevent decay), and the entire sample should then be placed in another, loosely folded plastic bag. Do not allow leaves, paper tags, or labels to contact with soil.

MUSHROOMS AND FRUITS: Wrap mushrooms or fruits in dry paper towels or newspaper and place in sturdy box to avoid crushing.

INSECTS: Send small insects in a small vial of alcohol (never in an envelope). Pack large insects, such as moths, in cotton.
Please **DO NOT SEND LIVE INSECTS**.

Turfgrass samples:

Plugs that are about 3-5" in diameter and deep enough to include the roots (usually about 3") are ideal. The best sample consists of a completely diseased plug, a healthy plug, and a plug from the transition zone between diseased and healthy turf.

Dutch elm disease testing and other vascular wilt testing:

Live, symptomatic branches that are at least 1" in diameter and 6-8" long should be submitted, with leaves attached.

Soil samples for nematode screen or Aphanomyces assay:

1. Use a soil probe to collect samples, 6-8 inches in depth.
2. Using a zigzag pattern, collect 10-20 soil cores per every 10 to 20 acres.
3. Collect cores from areas of similar soil type and crop history.
4. Dump cores from each 10-20 acre set into a bucket or tub and mix thoroughly.
5. For nematode screening, send 1 pint (2 cups) of mixed soil in a soil sampling bag or plastic zippered bag. For Aphanomyces indexing, send about 1 gallon (32 cups) of soil per 10-20 acre set. Label the bags with a permanent marker. Don't allow paper labels to come in contact with soil.
6. Store sample in a cool, dark place until shipped to the lab. Ship as soon as possible to: NDSU Plant Diagnostic Lab, 306 Walster Hall, Fargo ND 58102 (for non-USPS shippers); or NDSU Plant Diagnostic Lab, NDSU Dept 7660, PO Box 6050, Fargo ND 58108-6050 (for USPS).

Other tips:

- Send multiple specimens if available.
- Send sample as soon as possible after collecting.
- Don't add excess moisture to root ball. Use damp (not wet) paper toweling only.
- Avoid soil contact with foliage by wrapping root ball in separate plastic bag and tying off at base of stem.
- Use DRY paper towels on foliage to absorb moisture. Moisture on foliage causes rapid decay.
- Keep sample in a cool, dark place, such as a refrigerator, until ready to ship.
- Mail samples early in the week to avoid long delays over weekends at post offices. Samples mailed first class usually arrive at the lab in 1 day from most North Dakota and Red River Valley locations.